**ImmunoDOT™**

**Borrelia (Lyme) with Recombinant Protein**

**INTENDED USE**

The ImmunoDOT Borrelia (Lyme) test is an enzyme immunoassay (EIA) dot-blot test for the qualitative detection of borrelia antibodies. Both species specific (anti-P39) and related Borrelia burgdorferi antibodies (IgG, IgM, and IgA) are detected in serum as an aid in the diagnosis of Lyme disease.

**SUMMARY AND EXPLANATION**

Lyme borreliosis is reported to occur in North America, Europe, and Asia\(^{2,3,4,5}\). Lyme disease has been present in Europe for years, but was first recognized in the United States in 1975 in Lyme, Connecticut. The disease, transmitted through the bite of a tick infected with the spirochete, *Borrelia burgdorferi*, exhibits a variety of symptoms which may be confused with immune and inflammatory disorders. Inflammation around the tick bite eventually causing skin lesions, erythema chronicum migrans (ECM), is the first stage of disease. *B. burgdorferi* disease is also associated with neurologic or cardiac symptoms (stage 2) or arthritic (stage 3). A definitive distinction between stages is not always seen. In some cases, these secondary symptoms may occur even though the patient does not remember a tick bite or rash.

The criteria for the diagnosis of Lyme borreliosis are not clearly defined. Unless the typical ECM lesions are present, serological diagnosis is necessary to identify patients exposed to the agent. However, cross-reactions within the Borrelia genus and other cross-reactions (e.g., flagellin reactions with sprochetes and membrane reactions with bacterial membrane proteins) have limited the reliability of *B. burgdorferi* serology.

For confirmation of *B. burgdorferi* specific antibodies, ImmunoDOT Borrelia (Lyme) Test also measures the specific antibody response against the 39 kilodalton (P39) protein. Antibodies against P39, unlike the antibodies to flagellin which cross-react with other sprochetal flagellins, are specific to *Borrelia burgdorferi* and conserved among North American and European isolates.\(^{7}\) Additional unpublished studies have determined that other species of *Borrelia* (hermsii, parkeri, turicatae, and coniacea), *Leptospira* (icterohaemorrhagiae and canicola), and *Treponema pallidum and phagedenis* do not contain the *B. burgdorferi* P39 antigen. Because P39 protein is highly antigenic, but constitutes only a small fraction of the protein in the organism, it may not be detected in other assay systems (e.g., western blots, EIA, IFA, etc.) which are not enriched with P39 protein.

**ASSAY PRINCIPLE**

ImmunoDOT utilizes an enzyme-linked immunoassay (EIA) dot technique for the detection of antibodies. Various levels of antigens are dispensed as discrete dots onto a solid membrane. Because the *B. burgdorferi*-specific P39 protein represents a small fraction of borrelia protein, assay sensitivity is improved by addition of recombinant P39 antigen to the whole borrelial cell extract in the first Borrelia-reactive dot. To improve assay specificity serum is absorbed in a Diluent containing E. coli proteins. After specimen is absorbed with E.Coli extract, an assay strip is inserted, allowing patient antibodies reactive with the test antigen to bind to the strip's solid support membrane. In the second stage, the reaction is enhanced by removal of non-specifically bound materials. During the third stage, alkaline phosphatase-conjugated anti-human antibodies are allowed to react with bound patient antibodies. Finally, the strip is transferred to enzyme substrate reagent, which reacts with bound alkaline phosphatase to produce an easily seen, distinct dot.

**REAGENTS**

**Assay Strip** positive human antibody control, negative control, *B. burgdorferi* (strain B31) partially purified sonicate of whole organism blended with purified P39 recombinant protein, partially purified sonicate of whole organism, P39 recombinant protein, and flagellin extracted and purified from *B. burgdorferi* (strain B31).

**Diluent (#1)** buffered diluent (pH 6.2-7.6) containing solubilized *E. coli*, protein stabilizers with <0.1% NaNO<sub>2</sub>.

**Enhancer (#2)** sodium chloride with <0.1% NaNO<sub>2</sub>.

**Conjugate (#3)** alkaline phosphatase conjugated goat anti-human antibodies in buffered diluent (pH 6.2-8.5) with <0.1% NaNO<sub>2</sub>.

**Developer (#4)** 5-bromo-4-chloro-3-indolyl phosphate and p-nitro blue tetrazolium chloride in buffered diluent (pH 9-11) with <0.1% NaNO<sub>2</sub>.

**Warnings and Precautions**

**For In Vitro Diagnostic Use.** ImmunoDOT reagents have been optimized for use as a system. Do not substitute other manufacturers’ reagents or other ImmunoDOT Assay System reagents. Dilution or adulteration of these reagents may also affect the performance of the test. Do not use kit if evidence of microbial contamination (cloudiness) is present. Do not ingest reagents. Do not use any kits beyond the stated expiration date. Analytic quality deionized or distilled water must be used as Clarifier. Close adherence to the test procedure will assure optimal performance. Do not shorten or lengthen stated incubation times since this may result in poor assay performance.

Some assay components contain sodium azide which may react with lead and copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide build-up.

**Warning - Potential Biohazardous Material.** The antigens on the assay strip have been rendered noninfectious; nevertheless, caution should be exercised. Human sera used in the preparation of controls were tested and found non-reactive for hepatitis B surface antigen and for antibodies to HIV-1, HIV-2, and hepatitis C virus. Because no test method can offer complete assurance that infectious agents are absent, handle reagents and patient samples as if capable of transmitting infectious disease.

**Storage**

Store reagents and assay strips at 2-8°C. Reagents must be at room temperature (15-30°C) before use. Avoid contamination of reagents which may produce invalid results.

**SPECIMEN COLLECTION AND HANDLING**

ImmunoDOT Borrelia (Lyme) Test is performed on serum. Hemolyzed or lipemic serum has not been shown to be an acceptable specimen. Single specimens are used to assess exposure to *B. burgdorferi*. Two specimens collected at different times from the same individual are used to show seroconversion. The test requires approximately 10 µL of serum. Serum is collected according to standard practices and may be stored at 2-8°C for up to five days. Serum may be frozen below -20°C for extended periods.

**PROCEDURE**

**Materials Provided**

- Borrelia (Lyme) Assay Strips
- Reaction Vessels
- Diluent (#1)
- Package Insert
- Enhancer (#2)
- Conjugate (#3)
- Developer (#4)

**Materials Required But Not Provided**

- GenBio Workstation
- Specimen collection apparatus
- Timer
- Analytic quality distilled or deionized water to be used as Clarifier
- Absorbent toweling to blot dry the assay strip
- Positive control serum
- Pipets

**Set-Up**

1. Turn on Workstation and adjust to appropriate temperature if necessary. Refer to Workstation Instructions.
2. Remove 4 Reaction Vessels (per test) from the product box and insert into appropriate slots in Workstation. For the large Workstation, add water up to the fill line of the Clarifier Vessel.
provided. For the small Workstation, use an appropriate container and sufficient water to cover all reactive windows of the assay strip.

3. Place 2 mL Diluent (#1) in Reaction Vessel #1; 2 mL Enhancer (#2) in Reaction Vessel #2; 2 mL Conjugate (#3) in Reaction Vessel #3; and 2 mL Developer (#4) in Reaction Vessel #4.

4. Appropriately label the Assay Strips.

5. If the large Workstation is used, insert the label end of the assay strip into the Strip Holder, one per groove, taking care not to touch the assay windows.

**Assay Procedure**

1. Add 10 µL serum to Reaction Vessel #1 and incubate in the Workstation for 30-60 minutes.

2. Prewet Assay Strip by immersing in Clarifier for 30 - 60 seconds.

3. Using several (5-10) quick up and down motions with the Assay Strip, mix thoroughly in Reaction Vessel #1. Let stand for 5 minutes.

4. Remove Assay Strip from Reaction Vessel and swish in the Clarifier. Use a swift back and forth motion for 5-10 seconds allowing for optimal washing of the Assay Strip's membrane windows.

5. Place Assay Strip into Reaction Vessel #2. Mix thoroughly with several (5-10) quick up and down motions. Let stand for 5 minutes.

6. Remove Assay Strip from Reaction Vessel #2 and swish in Clarifier as described (Step #4).

7. Place Assay Strip into Reaction Vessel #3. Mix thoroughly with several (5-10) quick up and down motions. Let stand for 15 minutes.

8. Remove Assay Strip from Reaction Vessel #3 and swish in Clarifier as described (step #4). DO NOT remove the Assay Strip from the Clarifier.

9. Allow the Assay Strip to stand in the Clarifier for 5 minutes.

10. Remove Assay Strip from Clarifier and place into Reaction Vessel #4. Mix thoroughly with several (5-10) quick up and down motions. Let stand for 5 minutes.

11. Remove Assay Strip from Reaction Vessel #4 and swish in Clarifier as described (step #4).

12. Blot and allow Assay Strip to dry. It is imperative that tests of borderline specimens be interpreted after the Assay Strip has been allowed to dry.

**Reading the Assay Strip**

Positive A dot with an EASILY SEEN, distinct border is visible in the center of the window. The outer perimeter of the window must be white to pale gray.

Negative If no dot is seen or a dot is difficult to see, interpret it as negative.

Each window of the assay strip is interpreted independently. Reactions fall into three categories:

- **Nonreactive** Negative reaction
- **Weakly reactive** The dot is not easily seen and is interpreted as negative
- **Reactive** Positive reaction (“dot”)

Weakly reactive samples are sometimes seen and are negative reactions. Weak reactions may either indicate low level autoantibodies, nonspecific crossreacting antibodies which are found in normal subjects, or low level true reactions.

**INTERPRETATION**

The first dot, nearest the label is the positive reagent control. The second dot is the negative reagent control. The third dot measures both anti-P39 and anti-whole organism (Borrelia) responses. The fourth dot measures anti-whole organism. The fifth dot measures anti-P39. The bottom, or sixth dot, measures anti-flagellin. Because of the complex antibody cross-reactivities reported between B. burgdorferi antigens and other agents, the following reporting guideline is recommended:

1. Whenever both P39 and whole organism (dots 3,4, and 5) are reactive, the result is reported as:  
   "Specific B. burgdorferi detected"

2. Whenever all Borrelia dots are nonreactive, the result is reported as nonreactive for borrelia antibodies. Dot 3 (whole organism and P39) may exhibit a weak reaction while the other Borrelia dots are nonreactive. This result is also reported as:  
   "Borrelia antibody not detected"

3. All other outcomes are initially specified as reactive against the specific antigen (e.g., reactive against whole organism and nonreactive against P39, reactive against flagellin, etc.) or may simply be reported as:  
   "Borrelia antibody detected"

A dot with an EASILY SEEN, distinct border is visible in the center of the window. The outer perimeter of the window must be white to pale gray.

If no dot is seen or a dot is difficult to see, interpret it as negative.

Each window of the assay strip is interpreted independently. Reactions fall into three categories:

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If the reaction pattern obtained is not illustrated by one of the six examples above, contact GenBio Technical Service at (800) 288-4368.
If anti-P39 is absent, there are outcomes which may indicate Lyme borreliosis or may represent a false positive cross-reaction. In all cases an additional test to clarify the result is recommended before a definitive interpretation is reported. In cases in which clarification is not possible, another specimen collected two to six weeks later to measure an antibody response titer rise may provide additional information for diagnosis.

1. Flagellin: Because significant cross-reactivity between spirochetal and bacterial flagellin is possible, if the bottom dot (Dot 6), flagellin, is reactive and P39 (Dot 5) is nonreactive, an alternative test method such as western blot is recommended. If syphilis is suspected, a non-treponemal (e.g. RPR) test is recommended.

2. Whole Organism: Because of the known lack of specificity of the antibody response against whole borrelia organism, if P39 (Dot 5) is nonreactive, a second serology method using a different type of antigen preparation to clarify the reactivity is recommended. Although the western blot method has been shown to be no more reliable than any other method for Lyme borreliosis serological diagnosis, the method does provide additional information about antigen reactivity and may help clarify such indeterminate samples.

Quality Control

The top two membrane windows of the Assay Strip are reagent controls. The top window is a positive reagent control and must be positive for further interpretation. The next window is the reagent negative control and must be negative for further interpretation. Reagent controls assure that reagents are active and that the test has been performed properly. If either reagent control is invalid, the test must be repeated. The intensity of the positive control dot must not be used as a calibrator. Positive reactions in the antigen windows of the strip may be either darker or lighter than the positive control depending on the antibody titer.

A positive control serum (Product No. 3905), reactive in the total borrelia and P39 windows is available separately. Although other windows may be reactive, if the total borrelia or P39 windows are nonreactive, the control test is invalid. The performance of each kit lot may be confirmed upon receipt by running a determination using the positive control serum and obtaining a positive result. The control serum should be tested in accordance with laboratory guidelines.

The assay's reagent temperature is between 42-48°C. Due to heat transfer loss, the Workstation temperature is set higher. The appropriate Workstation temperature setting is listed in the Workstation's package insert. (Contact Technical Services for additional guidance if an alternate heat source is used.)

LIMITATIONS

- The diagnosis of Lyme disease should be based on interpretation of test results in combination with the patient's clinical signs and symptoms, other clinical and laboratory test results, and epidemiological data.
- Anti-P39 positive results should be interpreted in conjunction with other positive serological test results for Lyme disease.
- Negative results do not rule out the diagnosis of Lyme disease. Some patients may not produce significant humoral response to B. burgdorferi, early antibiotic therapy may suppress antibody response, P39 may not be expressed by all strains of B. burgdorferi, and/or detectable antibodies to P39 may not be produced in some Lyme disease patients, especially in early disease stages. Negative results in suspected early Lyme disease should be repeated in 4-6 weeks.
- The continued presence or absence of antibodies cannot be used to determine the success or failure of therapy.
- Testing should not be performed as a screening procedure for the general population. The predictive accuracy of a positive or negative serologic result depends on the pretest likelihood of Lyme disease being present. Testing should only be done when other laboratory tests and clinical evidence suggest the diagnosis of Lyme disease.
- A single positive result only indicates previous immunologic exposure; level of antibody response is unreliable in determining active infection or disease stage.
- Whole organism or flagellin positive results should be interpreted with caution. Sera from patients with other spirochetal diseases (syphilis, yaws, pinta, leprospirosis, relapsing fever, peridontal disease, etc.) may also give positive results. If syphilis is suspected, a non-treponemal (e.g., RPR) test may be used to rule out this cause.

- Patients with connective tissue autoimmune diseases (rheumatoid arthritis, ANA, SLE, etc.) may have antibodies which cross-react with whole organism antigen(s).
- Patients with other bacterial and viral infections such as Rocky Mountain Spotted Fever, EBV, CMV, and HIV may also have antibodies which cross-react with B. burgdorferi antigens.

EXPECTED RESULTS

In general, three types (stages) of Lyme disease are recognized: erythema chronicum migrans (ECM), neurologic, and arthritic. Antibody levels are generally low or absent during early (ECM) infection. Most symptomatic patients will have either no antibody or highly cross-reactive antibody during the first 1-2 weeks after tick bite and the antibody titer will rise and become more specific with time. Highest antibody levels are seen in chronic arthritis subjects.

The number of antibody positive subjects in a population depends on several factors: 1) prevalence of the causative agent, 2) assay used to detect antibody, and 3) clinical screening criteria to select tested subjects. Because early assays lacked accuracy, the number of antibody positive subjects in a population at present (1991) is highly dependent on the assay used. Whenever a suitably accurate test is used, few positives should be detected in a randomly screened population in an endemic area. On the other hand, if patients with typical ECM signs in an endemic region are tested, many positive results are expected.

Disagreement between assays which do not use an absorbent and those assays like ImmunoDOT which do use an absorbent are expected. Fawcett has shown that assays using an absorbent were equally sensitive to those without an absorbent and that the absorbed assays were significantly more specific. In the course of primary disease, the highly specific anti-P39 antibody may appear after earlier, non-specific antibody. Anti-P39 was positive in all Stage 3 (arthritic) cases tested.

PERFORMANCE CHARACTERISTICS

Reproducibility

Because ImmunoDOT products are discrete, qualitative tests, within assay reproducibility is not applicable. Between assays reproducibility is demonstrated by testing the same series of positive samples. In all cases, the samples interpreted as positive in the ImmunoDOT product are repeatedly positive (six replicates) in the ImmunoDOT product.

Correlation

The specificity of ImmunoDOT Borrelia (Lyme) Test using 196 sera from asymptomatic blood donors collected from areas other than the upper Midwestern and Northeastern United States (hyperendemic regions) is 100% specific.

The specificity was also determined using 168 sera from non-Lyme, symptomatic patients (Table 1), with autoimmune disease, pneumonia, mononucleosis, lymphadnopathy, rheumatoid arthritis, syphilis, and AIDS. No samples were positive for anti-P39 (100% specificity); however, all syphilis samples reacted with the dots containing borrelia antigens other than P39.

Table 1: SPECIFICITY-Lyme Disease Negative Patients

<table>
<thead>
<tr>
<th>Sample</th>
<th>Nonreactive</th>
<th>Borrelia +</th>
<th>B.burgdorferi +</th>
</tr>
</thead>
<tbody>
<tr>
<td>SLE</td>
<td>5</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>JRE</td>
<td>11</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>ANA+</td>
<td>36</td>
<td>2</td>
<td>0</td>
</tr>
<tr>
<td>CMV IgM+</td>
<td>25</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Heterophile+</td>
<td>24</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>RF +</td>
<td>24</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>HIV +</td>
<td>15</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>RMSF+</td>
<td>12</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Syphilis+</td>
<td>0</td>
<td>12</td>
<td>0</td>
</tr>
</tbody>
</table>

Sera from eighty-four patients at three sites diagnosed with Lyme borreliosis were used to measure assay sensitivity. Diagnoses were based on epidemiological, clinical, and serological criteria. These studies are presented in Table 2.
Table 2: SENSITIVITY-Diagnosed Lyme Disease Patients

<table>
<thead>
<tr>
<th>Sample</th>
<th>Nonreactive</th>
<th>Borrelia +</th>
<th>B. burgdorferi +</th>
</tr>
</thead>
<tbody>
<tr>
<td>Site 1</td>
<td>0</td>
<td>45</td>
<td>41</td>
</tr>
<tr>
<td>Site 2</td>
<td>1</td>
<td>11</td>
<td>11</td>
</tr>
<tr>
<td>Site 3-Early (ECM)*</td>
<td>18</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>Site 3-Late</td>
<td>0</td>
<td>5</td>
<td>5</td>
</tr>
<tr>
<td>Totals</td>
<td>19</td>
<td>65</td>
<td>57</td>
</tr>
</tbody>
</table>

* All 22 samples were EIA and western blot negative.

Assay sensitivity for the samples which were not classified as to stage of disease was 98% (56/57). The one nonreactive sample was also nonreactive in other EIA tests, IFA, and western blot. Fifty-two (91%) were anti-P39 reactive. Consistent with expectations that antibody is either absent or at low titer in early cases, four of 22 sera (18%) from early Lyme cases with ECM, but nonreactive in EIA and western blot analysis, were ImmunoDOT reactive for borrelia antibody. None were anti-P39 reactive. All sera from the five late stage cases were positive.

Assay specificity is 100% in asymptomatic normals. Specificity in Lyme disease negative patients other than those with syphilis is 97%. None were anti-P39 reactive. All 12 syphilis subjects contained anti-borrelia, but none were anti-P39 reactive.

Eighty-seven samples were evaluated at Site 1 using the ImmunoDOT and anticomplement immunofluorescence (ACIF) methods. The comparative results are shown in Table 3.

Table 3: Correlation to ACIF

<table>
<thead>
<tr>
<th>ACIF</th>
<th>Nonreactive</th>
<th>Borrelia +</th>
<th>B. burgdorferi +</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reactive</td>
<td>0</td>
<td>45</td>
<td>41</td>
</tr>
<tr>
<td>Nonreactive</td>
<td>38</td>
<td>4</td>
<td>0</td>
</tr>
</tbody>
</table>

Twenty-four proficiency samples from a state public health laboratory and a national proficiency program were also used to evaluate ImmunoDOT performance. The expected results (reactive or nonreactive) were based on consensus. That is, the samples were classified as reactive if almost all reported results were reactive, and vice-versa for nonreactive specimens. ImmunoDOT detected 14 out of 14 nonreactive samples correctly, and identified 10 of 10 reactive samples as borrelia antibody reactive, but only identified five of the ten as anti-P39 reactive. These results indicate that the five samples without anti-P39 are either cross-reactive or lack specific anti-P39, presumably due to the early stage of disease.

BIBLIOGRAPHY


8. Centers for Disease Control/National Institutes of Health Manual Biosafety in Microbiological and Biomedical Laboratories (1983)
